NRT Quick Reference Guide:  
Francisella tularensis (Causes the disease Tularemia)  

For reference, please see "Key References Cited/Used in National Response Team (NRT) Quick Reference Guides (QRGs) for Bacterial 2011 Revision.
QRGs are intended for Federal On-Scene Coordinators (OSCs) and Remedial Program Managers (RPMs)."

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**Agent Characteristics**

| Bio-Safety Level: | 3 |
| CDC Class: | A |
| HHS Select Agent: | Yes |
| Incubation Period: | 1-10 days, up to 21, but typically 3-5 days. |
| Duration of Illness: | 2 or more weeks, depending on how quickly treatment is provided |
| Person-to-Person Transmission: | No |

**Release Scenarios**

- **Air:** F. tularensis can be made for dry or wet aerosol dispersion. Re-aerosolization is considered with weaponized F. tularensis, though it is thought to be unlikely. F. tularensis has been weaponized for dry & wet aerosol dispersion. The area of initial release might be difficult to identify because symptoms may take days to appear.
- **Soil:** F. tularensis exists naturally (endemic in North America & Eurasia) & can contaminate hay, soil & can live for weeks in cold, moist conditions.
- **Surfaces:** Unknown, but could be used as a transmission point.
- **Water:** Possible pathway for weaponized strain, and natural strain outbreaks have also occurred.

**Onset**

- 3-5 days after aerosolized release, may present within 1-10 days & up to 21 days post exposure.

**Signs & Symptoms**

- **General:** The primary clinical forms of tularemia will vary in severity & presentation depending on virulence of the infecting organism, dose, & site of infection. Tularemia can be fatal without treatment & is an incapacitating disease in non-fatal cases.
- **Inhalation:** Inhalation of F. tularensis can produce flu-like symptoms progressing to pneumonia, ulcers of the mouth, chest pain, breathing difficulty, bloody sputum, & respiratory failure.
- **Skin:** Cutaneous infection through abraded or cracked skin resulting in ulcers at the infection site & swollen lymph nodes. Cutaneous infections also result from bites from ticks and biting insects that have fed on infected animals or carcasses.
- **Ingestion:** Eating/Drinking contaminated food/water can cause oropharyngeal tularemia; typhoidal and gastrointestinal presentations may also occur however a large numbers of organisms are required (10ES to 10GB).

**Effect Levels**

- **Infectivity:** The organism is highly infectious by the inhalation route. Long term immunity usually follows recovery from tularemia. However, re-infection has been reported.
- **Lethality:** Less than 7% for pneumatic and typhoidal. If appropriate antibiotic therapy is provided early. The other forms are usually not fatal.

**Concerns**

- Check with the Health & Safety Officer regarding PPE, Medical Surveillance, & Health & Safety Plan (HASP). Level of PPE may vary depending upon the incident & site-specific circumstances. The PPE levels listed are general suggestions only & are appropriate only for F. tularensis; they may not provide protection for some decon & other chemicals that workers may be exposed to during response/recovery operations. For decon of workers, use warm soapy water, taking care to avoid abraded the skin.

**Medical**

- **Baseline:** Annual physical & respiratory function exams. THERE IS NO FDA APPROVED HUMAN VACCINE FOR TULAREMIA.

**PPE**

- **Emergency Response to a Suspected Biological Incident:** Possible PPE Levels for emergency responders is based on scenario risks from highest level of protection to lowest: 1) Pressure-demand Self Contained Breathing Apparatus (SCBA) with Level A protective suit, when: a) Event is uncontrolled, b) The type(s) of airborne agent(s) is unknown, c) The dissemination method is unknown, d) Dissemination via an aerosol-generating device has stopped, but there is no information on the duration of dissemination, or what the exposure concentration may be. 2) Pressure-demand SCBA with Level B protective suit, when: a) The suspected biological agent is no longer being released, b) Other conditions may present a splash hazard. 3) Full-facepiece respirator with P100 filter or PAPR with HEPA filters, when: An aerosol-generating device was not used to create high airborne concentrations. 4) Disposable hooded coveralls, gloves, & foot coverings, when: Dissemination was a by a letter, package, or other material that can be bagged, contained, etc.

**Other Workers:** PPE recommendations for workers other than emergency responders must be developed in the HASP for the specific scenario. PPE recommendations will vary by job type (e.g., cleanup, decon, etc.), type of exposure (e.g., airborne or surface/liquid/solid soil), & any other site hazards (e.g., chemical, physical, etc.).

**Field Detection**

- Fixed Aerosol Monitoring: An aerosol release of F. tularensis may be detected using air samples & PCR. Results may be delayed as much as 2 days from time of release. In the absence of reliable detection, a F. tularensis release will only be confirmed once patients present with symptoms and are diagnosed or animal die-off from Tularemia is confirmed. Consult EPA/HAO-EOC at 202-564-3850 for more information.

**Portable Aerosol Monitoring:** Portable aerosal monitoring may use dry or wet sampling methods. Dry sampling (useful only for molecular analyses) includes gelatin, cellulose acetate & Teflon methods. Wet sampling methods include liquid impingers (low flow) & impactors (single or stage). Refer to the manufacturer’s aseptic sampling methods, flow rates, & sampling times. Ensure that the appropriate pump is used for the selected sampling method.

**Sampling**

- **Concerns:** BEFORE OBTAINING SAMPLES: Identify sample transportation requirements; Contact EPA/HAO-EOC (202-564-3850) for ERLN contract laboratories able to analyze these types of samples; Clearly identify & coordinate with the laboratory to be used since most labs cannot analyze all types of media (e.g., wipes, swabs, and HEPA vacuum samples); Coordinate with the sample disposal facility for acceptance criteria (i.e., sample decon requirements); Coordinate with investigative units (EPA-CID & FBI) to ensure sample chain-of-custody is maintained between the groups. Note: Detection/analytical equipment & sampling techniques will be highly site-specific & depend on: 1) The characteristics of the agent; 2) The type of contaminated surfaces (e.g., porous v. nonporous); 3) The phases/purposes of sampling (initial ID v. post-decon sampling); 4) The way in which samples are handled so as not to adversely affect viability; 5) Transportation regulations 6) The acceptance criteria of the analytical laboratory & 7) The sample decon requirements for the waste disposal facilities to be used. See LABORATORY ANALYSIS, below.

**CAUTION:** ONLY MANUFACTURER CERTIFIED HEPA VACUUM EQUIPMENT SHOULD BE USED. A site-specific sampling plan should be reviewed & approved by appropriate Subject Matter Experts &/or through ICS channels.

**Sampling Location Plans:** If release was limited to a small area due to opening a letter or container, start with an area thought to be free of contamination & work in concentric circles towards the initial point of contamination. Consider being concerned about other contaminated areas due to foot traffic/ventilation systems (e.g., elevator buttons, mail, corners of hallways, baseboards, light switches, door knobs, etc.). Based on site characteristics & laboratory capacity, the sampling plan may be judgmental, probabilistic, or a combination thereof.

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Consult EPA/HQ-EOC at 202-564-3850 for Environmental Response Laboratory Network (a.k.a. ERLN laboratory) contact information for personnel who can explain/describe the sampling procedure most compatible with their current analytical procedure.

**Types of Samples:** Air, water, soil, surfaces, agriculture & wildlife

**Materials:** While F. tularensis DNA can be detected long after the bacteria themselves have perished & might be of forensic interest, the presence of the DNA says little about the potential human risk in the days following a release.

**Air:** Dry sampling (useful only for molecular analyses) includes gelatin, cellulose acetate & Teflon methods. Wet sampling methods include liquid impingers (low flow) & impactors (single or six stage). Refer to the manufacturer’s aspecfic sampling methods, flow rates, & sampling times. Ensure that the appropriate pump is used for the selected sampling method.

**Water:** Since F. tularensis can persist in water, any potable water source should be sampled. If the potable water is chlorinated, the chlorine needs to be neutralized immediately with a sodium thiosulfate or other neutralizer at the concentration specified by the analytical laboratory prior to shipment. As chlorine levels can vary substantially throughout a drinking water system, it is not always appropriate to assume that a sample is chlorinated based solely on a description of the water treatment processes in use.

**Soil:** For the localized areas where soil deposition of the agent is suspected (i.e., aerosol or liquid droplets), a surface soil sample from a depth of less than 1 inch (2.54 cm) should be obtained from a non-vegetated area.

**Surfaces:** 1) Wipe & Swab Sampling (for non-porous surfaces): Sterile macrofoam swabs moistened with 1X phosphate-buffered saline supplemented with 0.01% Tween-20 (PBST). If this solution is not available, use sterile de-ionized water (DI). Do NOT use dry wipes or swabs. 2) HEPA Vacuum Sampling (for both porous & non-porous surfaces): collect samples in a HEPA sock designed to fit into an inlet nozzle of a manufacturer certified HEPA vacuum cleaner. Good for screening & determining the extent & location of contamination in large areas.

**Agreements:** Upon confirmation of an outbreak, these agencies are notified immediately since tularemia is a zoonotic vector borne disease; USDA at 202-720-5711 & National Center for Emerging and Zoonotic Infectious Diseases at 800-232-4636 (after hours call the Directors Emergency Operations Center at 770-488-7100).

**Samples that test for Re-aerosolization:** 1) Wipe sampling of the air duct system (filters, areas of particulate deposition) if exposure occurred indoors. 2) Air Samplers & Single Stage Impactors with settle plates for capturing airborne particulates of respirable size (1-5 microns) on a series of agar plates. Agar plates are then sent to laboratory for culture analysis.

**Sample Packaging & Shipping:** The packaging & shipping of samples is subject to strict regulations established by DOT, CDC, USPS, OSHA, & IATA. Contact the sample-receiving laboratory to determine if they have additional packaging, shipping or labeling requirements (e.g., DO NOT X-RAY). Samples should be packaged in an air-tight container & kept at temperatures of 40-50°F (4-10°C). Ensure samples are not placed directly on the ice used for cooling the shipping container.

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**Laboratory Information:** F. tularensis may be highly aerosolizable. Contact EPA/HQ-EOC at 202-564-3850 for ERLN laboratories that can analyze these types of samples.

**Laboratory Analysis:** Pilot study to determine if viable F. tularensis is present in the sample. Note: The selected laboratory may use a tiered approach. If a tiered approach is used, the initial analysis may only determine if select/compatible components of the bacterium are present in the sample (e.g., presence or absence). It may take additional time (up to weeks depending on the laboratory) to determine if the bacterium are viable & still able to cause adverse effects.

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**Decon Planning:** Site-specific decon/cleanup plan should be developed & approved by all necessary organizations/SMEs via ICS channels. Responders should develop a plan that takes into account: 1) Nature of contamination including purity, physical properties, how it entered the facility, etc.; 2) Extent of contamination, including the amount & possible pathways that have spread the agent. It is advisable to isolate the contaminated area, & 3) Objectives of decon, including decon of critical items for re-use & the treatment, removal, or packaging of other items for disposal. Note: Crisis exemptions from EPA’s Office of Pesticide Programs might be necessary depending on decontaminating agents used.

**CAUTION:** DECON SOLUTIONS SHOULD NOT BE DEPLOYED AS A SPRAY WHENEVER POSSIBLE.

**Decon Methods:** Decon decisions will be site & situation specific but due to re-aerosolization concerns, under NO circumstances should a non-HEPA vacuum cleaner or a broom be used. EPA’s National Decon Team (800-329-1841) can provide specific decontamination parameters & requirements for using readily available commercial items such as household bleach.

**Methods used on surfaces:** 1) Source reduction steps, including HEPA vacuuming; 2) Liquid antimicrobial products such as pH-amended bleach (mixture of 1 part household bleach (5.25% to 6.0%) to 1 part white vinegar to 8 parts water, is recommended). This product affects surfaces differently in terms of corrosiveness, staining, & residue. The product will be most efficient a) at higher temperatures (i.e., >70°F or 21°C) b) when plain bleach (e.g., no added fragrance) is used to make the pH-amended bleach solution, c) when pH is < 7. d) when presence of other surface contaminants is minimal, & e) when surfaces remain wet with amended bleach solution for 60 minutes. Note: Store-bought bleach does degrade with time – check the expiration date. Alternate antimicrobial products include: chlorine dioxide, hydrogen peroxide, & peroxyacetic acid. Fumigation: Uses gas or vapor to decontaminate facilities in which there is contamination, re-aerosolization, or if decontamination of limited access areas is required (e.g. HVAC systems). Fumigants: chlorine dioxide, & vaporized hydrogen peroxide. Prior to use, the fumigant’s compatibility with materials, penetration capacity, method of removal at the end of fumigation, as well as it’s physical, chemical, & toxicological properties should be taken into account. Each chemical has a specified range for process variables (e.g., temperature, relative humidity, concentration & contact time) that must be followed. Other Decon: 1) Ethylene oxide sterilization is used to decontaminate items in an off-site sterilization chamber. 2) Irradiation uses cobalt-60 & electron beam technology to destroy agents at off-site locations. This procedure may destroy magnetic media. Irradiation & chemical sterilization may be useful in decontaminating items that are intended to be returned to owners.

**Verification of Decon:** Site & situation specific. Please contact ERT (732-321-6660) and NDT (800-329-1841) for further assistance.

**CAUTION:** Hazardous waste transportation & disposal are regulated federally; however, more stringent regulations may exist under state authority. These regulations differ from state-to-state. Detailed state regulations can be found at: http://www.envcap.org.

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